



RESPIRATORY SYNCYTIAL VIRUS (RSV) AEROSOL EFFICACY TESTING

PROJECT: GPS AEROSOL RSV

TECHNOLOGY: Needle Point Bipolar Ionization

DEVICE: GPS-FC48-AC™

CAP LIC NO: 8860298

CLIA LIC NO: 05D0955926

STATE ID: CLF 00324630

CHALLENGE ORGANISM(S):

RESPIRATORY SYNCYTIAL VIRUS (RSV)

Dana Yee, M.D.

Medical Director

Study Completion Date

7/26/21

Testing Facility

Innovative Bioanalysis, Inc.

3188 Airway Ave Suite D

Costa Mesa CA, 92626

www.InnovativeBioanalysis.com

Email: info@innovativeBioanalysis.com

Laboratory Project Number

1034-R



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Efficacy Study Summary

Study Title	RESPIRATORY SYNCYTIAL VIRUS (RSV) AEROSOL EFFICACY TESTING
Laboratory Project #	1034-R
Guideline:	No standard exists; GCLP and modified ISO standards were used.
Testing Facility	Innovative Bioanalysis, Inc.
GLP Compliance	All internal SOPs and processes follow GCLP guidelines and recommendations.
Test Substance	Respiratory Syncytial Virus (RSV)
Description	The GPS-FC48-AC™ device housing NPBI™ technology is commercially available and designed to be installed in the ductwork of an HVAC system to reduce the concentration of certain bacteria and viruses while operational. Testing was conducted on the device to evaluate the effectiveness of the NPBI™ technology in reducing aerosolized Respiratory Syncytial Virus (RSV).
Test Conditions	The test was conducted in a airtight 20'x8'x8' chamber with a redundant negative pressure system connected to HEPA filters and an in-duct UV-C system. The temperature during testing was $72 \pm 2^\circ\text{F}$, with a relative humidity of 46%. Aerosolization was generated by filling a nebulizer with a suspension media containing an RSV concentration of 3.26×10^6 TCID50/mL. Air samples were collected after 0, 15, 30, 45, and 60 minutes of exposure to the operating device.
Test Results	The GPS-FC48-AC™ device housing NPBI™ technology decreased the concentration of Respiratory Syncytial Virus from a starting concentration of 3.26×10^6 TCID50/mL to 9.65×10^4 TCID50/mL after 60 minutes of the GPS FC48-AC™ in operation. Ion concentrations were measured in the chamber during a dry run test prior to viral challenges with an average of 22,000 negative ions per cm^3 .
Control Results	Through the duration of testing, RSV decreased from 3.26×10^6 TCID50/mL to 1.83×10^6 TCID50/mL. The results for the controls were plotted to show a natural rate of loss over 60 minutes and were used to assess the NPBI™ technology's ability to reduce RSV in air.
Conclusion	The NPBI™ technology exhibited the ability to neutralize aerosolized RSV faster than the natural viability loss observed in the control at each time point. A 97.04% reduction in viral concentrations in the air after 60 minutes of exposure was achieved.



Study Report

Study Title: RESPIRATORY SYNCYTIAL VIRUS (RSV) AEROSOL EFFICACY TESTING

Sponsor: Global Plasma Solutions

Test Facility: Innovative Bioanalysis, Inc. 3188 Airway Ave Suite D, Costa Mesa CA, 92626

Technology Tested: NPBI™

Device Tested: GPS-FC48-AC™

Study Report Date: 07/28/2021

Experimental State Date: 04/12/2021

Experimental End Date: 04/16/2021

Study Completion Date: 07/26/2021

Study Objective:

An ionization unit, GPS-FC48-AC™ containing NPBI™ technology, was provided by Global Plasma Solutions for testing to evaluate the efficacy against an aerosolized virus, Respiratory Syncytial Virus (RSV).

Test Method:

Bioaerosol Generation:

The nebulizer was filled with a 3.26×10^6 TCID50 per mL suspension of Respiratory Syncytial Virus and nebulized at a flow rate of 1mL/min with untreated local atmospheric air. Upon each completion, the nebulizer's remaining viral stock volume was weighed to confirm that roughly the same amount was nebulized during each test run. Bioaerosol procedures for the controls and viral challenges were performed in the same manner with corresponding time points and collection rates.

Bioaerosol Sampling:

Four probes connected to calibrated Gilian 10i vacuum devices set at a standard flow of 5.02L/min with a 0.20% tolerance were inspected for functionality before being used. Sample collection volumes were set to 10-minute draws per time point. The air sampler operated in conjunction with a removable sealed cassette and manually removed after each sampling time point. Cassettes had a delicate internal filtration disc to collect viral samples, which was moistened with a viral suspension media to aid in the collection. Filtration discs from Zefon International, Lot# 24320, were used for testing.

Test System Strains: Respiratory Syncytial Virus (RSV)

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Study Materials and Equipment:

Equipment Overview: The GPS-FC48-AC™ device housing NPBI™ technology arrived at the laboratory pre-packaged from the manufacturer and was inspected for damage upon arrival. Before starting the challenge, the GPS-FC48-AC™ was operated for 1 hour in a dry run to confirm correct operations.

MANUFACTURER: Global Plasma Solutions

MODEL: GPS-FC48-AC™

SERIAL #: N/A



Testing Layout:

Testing was conducted in a 20'x8'x8' sealed chamber per Biosafety Level 3 (BSL3) standards. The overall dimensions of the test chamber provided a displacement volume of 1,280 cubic feet and approximately 36,245.56 liters of air. The device was placed in the room's centerline, mounted on a movable scaffolding against the wall at an elevated position six feet above the ground, depicted in Figure 1. A variable-speed fan was placed behind the GPS-FC48-AC™ to create the necessary airflow to produce the required concentration of negative and positive ions. During testing, ion measurements were taken to confirm consistent readings, as shown in Figure 2 & 3.

At each chamber corner, low-volume mixing fans moving at approximately 120 CFM were positioned at 45-degree angles to ensure homogenous mixing of bioaerosol concentrations when nebulized into the chamber. For air sample testing, the room was equipped with four probes that were positioned along the centerline of the room and protruded down from the ceiling 24". A nebulizing port connected to a programmable compressor system was located in the center of the 20' wall protruding 24" from the wall. Due to the nature of ions, there were fluctuations of concentrations around the entire room. Ion readings were taken from multiple points in the room before aerosol testing, as shown in Figure 2. The chamber was visually inspected, pressure tested, and all internal lab systems and equipment were reviewed before testing.

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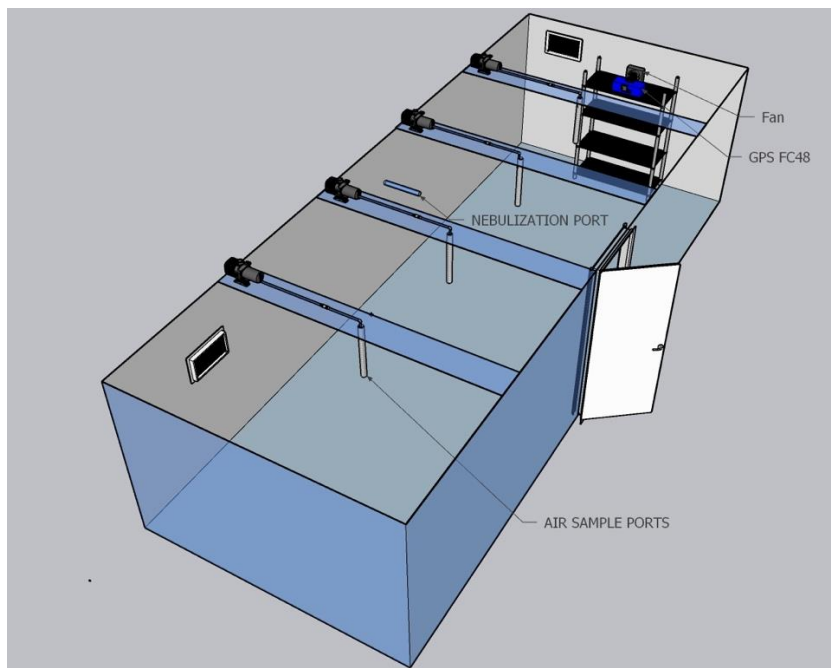


Figure 1. Room layout for the control and experimental trials.

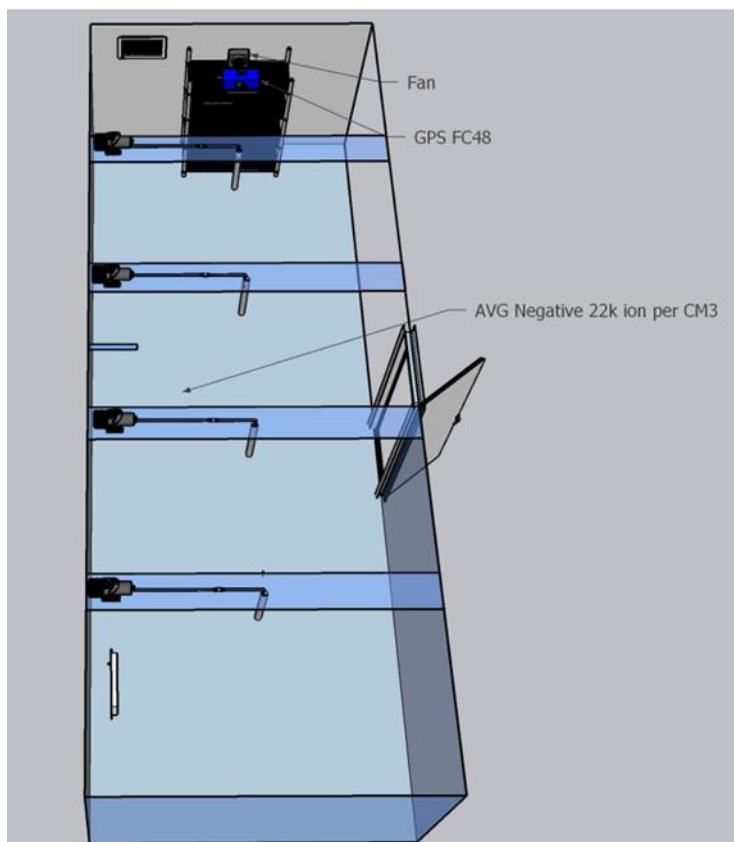


Figure 2. Overhead view of the dry run ion concentration observations.

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Test Method:

Exposure Conditions:

1. The temperature during all test runs was approximately 72°F ±2°F with a relative humidity of 47%.
2. Testing time points were as follows, with T equal to minutes: T-0, T-15, T-30, T-45, and T-60.

Nebulization:

1. The testing area was decontaminated and prepped per internal procedures before the initial control test and following each trial run.
2. An RSV stock of 3.26×10^6 TCID₅₀/mL in FBS media was nebulized into the sealed environment via the dissemination port.
3. After nebulization, the GPS-FC48-AC™ device housing NPBI™ technology was turned on via remote control.
4. Air sampling collection occurred after nebulization ceased for the challenges and control test.
5. After each run, sample cassettes were manually removed from the collection system and taken to an adjacent biosafety cabinet to be pooled.

Post Decontamination:

After each viral challenge test, the UV system inside the testing chamber was activated for 30 minutes. After 30 minutes of UV exposure, there was a 30-minute air purge through the air filtration system. All test equipment was cleaned at the end of each day with a 70% alcohol solution. Collection lines were soaked in a bleach bath mixture for 30 minutes then rinsed repeatedly with DI water. The nebulizer and vacuum collection pumps were decontaminated with hydrogen peroxide mixtures.

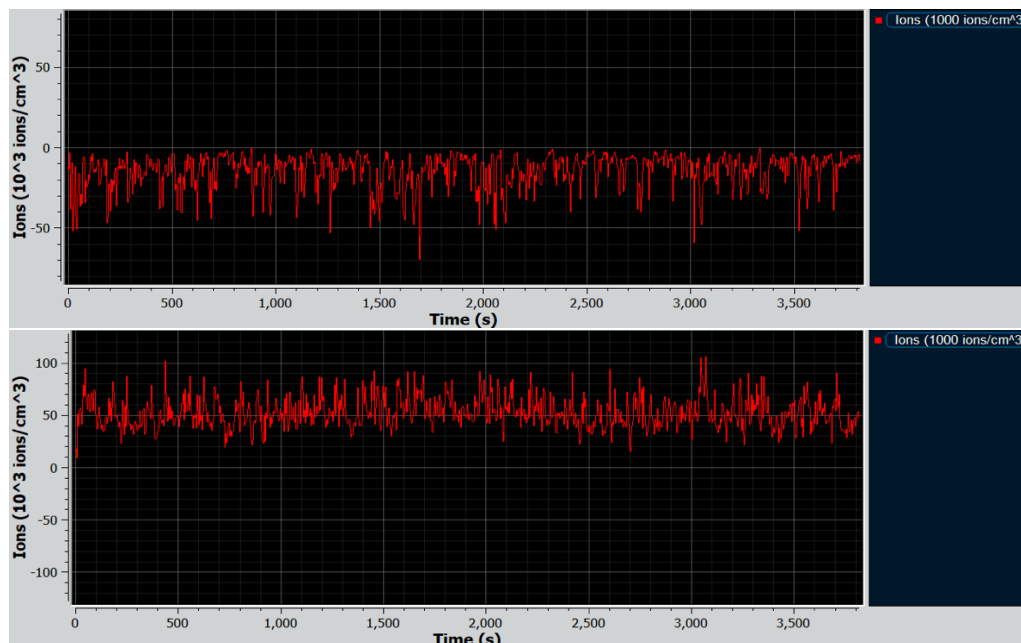


Figure 3. Device ion concentration recordings while in operation during testing.



Preparation of The Pathogen

Viral Stock: Human Respiratory Syncytial Virus (NR-28525, Lot #: 60109225)

Test	Specifications	Results
Identification by Infectivity in HE-p2 Cells	Cell rounding and sloughing	Cell rounding and sloughing
Sequencing of Species-Specific Region (822 nucleotides)	Consistent with human respiratory syncytial virus, A2001/2-20	Consistent with human respiratory syncytial virus, A2001/2-20 GenBank: JX069798
Titer by TCID50 in Hep-2 Cells	Report Results	2.8 X 10 ⁸ TCID50 per mL
Sterility (21-Day Incubation)		
Harpos HTYE Broth, aerobic	No Growth	No Growth
Trypticase Soy Broth, aerobic	No Growth	No Growth
Sabourad Broth, aerobic	No Growth	No Growth
Sheep Blood Agar, aerobic	No Growth	No Growth
Sheep Blood Agar, anaerobic	No Growth	No Growth
Thioglycollate Broth, anaerobic	No Growth	No Growth
DMEM with 10% FBS	No Growth	No Growth
Mycoplasma Contamination		
Agar and Broth Culture	None Detected	None Detected
DNA Detection by PCR of extracted test article nucleic acid	None Detected	None Detected

*The viral titer listed in the Certificate of Analysis represents the titer provided by BEI Resources. These viruses are grown on 10- to 11-day-old SPF Embryonated Chicken Eggs either in-house or at a partner lab to the concentrations listed within the experiment design.

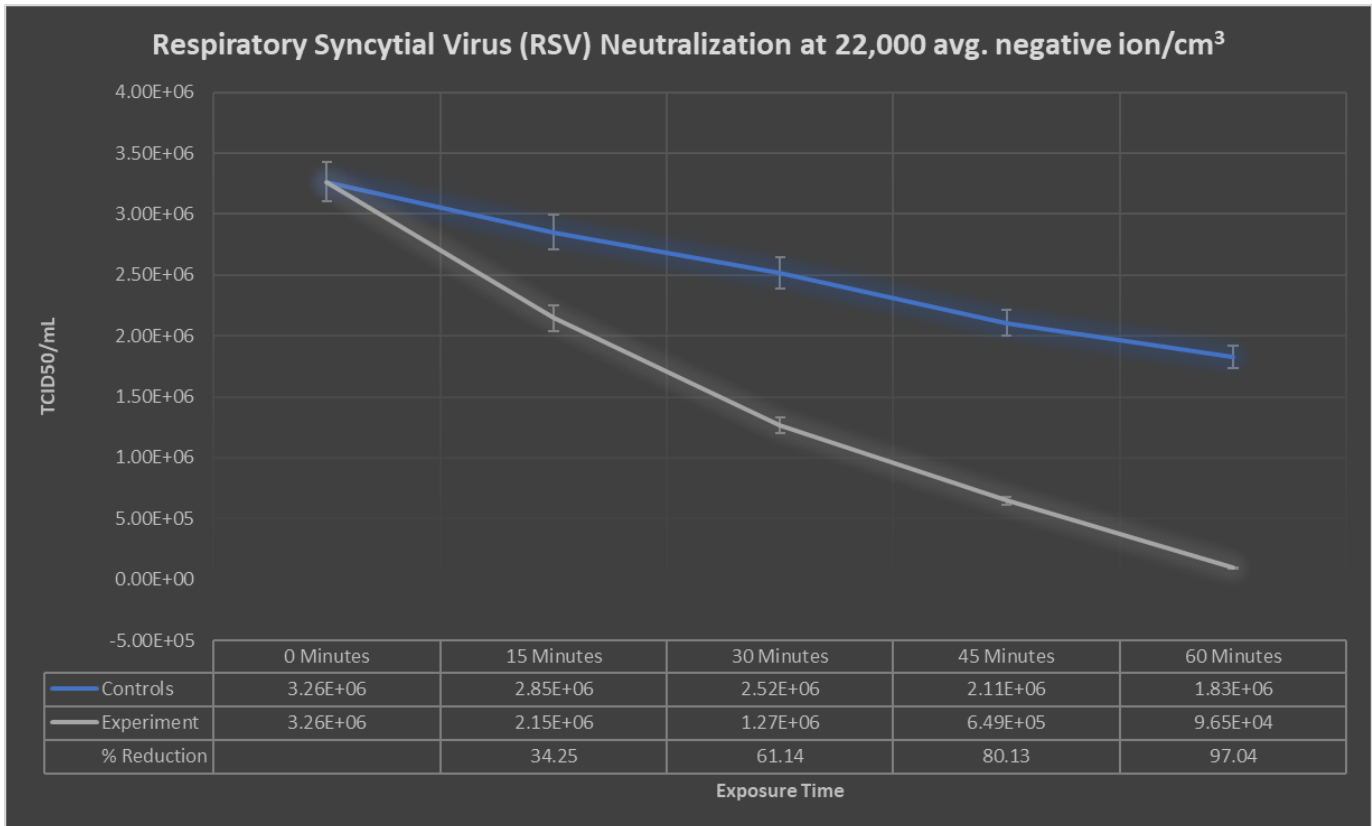
Control Protocol

To accurately assess the GPS-FC48-AC™ device housing NPBI™ technology a control was conducted without the device operating in the testing chamber. The collection was taken at corresponding time points used for the challenge trial, in the same manner, to serve as a comparative baseline to assess aerosolized viral reduction when the device was operating.

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Study Results



Conclusion:

The GPS-FC48-AC™ device housing NPBI™ technology displayed a significant impact against RSV under controlled settings. The equipment reduced active RSV by 97.04% in the air after 60 minutes of operation. Ion concentrations were measured in the chamber during a dry run test prior to viral challenges with an average of 22,000 negative ions per cm³. Overall, the device showed the capability of lowering aerosolized viral pathogen RSV at a higher rate than the natural loss observed in control.

Considerations:

When working with microorganisms and collecting said microorganisms, some variables cannot be fully accounted for, namely, placement of microorganisms, collection volume, collection points, surface saturation, microorganism destruction on collection, and possibly others. Every effort was made to address these constraints with the design and execution of the trials. And these efforts are reflected in the meaningful recovery of microorganisms in the control test.

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DocuSigned by:

Dana Yee

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8/24/2021

Dana Yee M.D

Date

Clinical Pathologist and Medical Director, Innovative Bioanalysis, Inc.

DocuSigned by:

Sam Kabbani

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8/24/2021

Sam Kabbani, MS, BS, MT(ASCP), CLS

Date

Chief Scientific Officer, Innovative Bioanalysis, Inc.

DocuSigned by:

Albert Brockman

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8/24/2021

Albert Brockman

Date

Chief Biosafety Officer, Innovative Bioanalysis, Inc.

DocuSigned by:

Kevin Noble

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8/24/2021

Kevin Noble

Date

Laboratory Director, Innovative Bioanalysis, Inc.

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